

Development and Formulation of Probiotic Using Fermentation Method

Aakansha Ravindra Patil^{1*}, Rutik Asaram Hajare², Abhay Bajrang Jadhav³, Kalpita Manik Patil⁴, Kumudini Kisan Pandharkar⁵

¹Doctorate Of Pharmacy, Bharati Vidyapeeth Poona College of Pharmacy
ap826893@gmail.com

²Bachelor Of Pharmacy, Indala Institute of Pharmacy
rutikhajare26522@gmail.com

³Bachelor Of Pharmacy, Indala Institute of Pharmacy
abhayjadhav6540@gmail.com

⁴MSc. Medical Genetic, MGM School of Biomedical Sciences
kalpitapatil01@gmail.com

⁵MSc. Medical Genetic, MGM School of Biomedical Sciences
pandharkarkumudini@gmail.com

Abstract

The present study focuses on the development and formulation of a probiotic using a fermentation-based approach with blueberry (*Vaccinium corymbosum*) and black rice (*Oryza sativa L. indica*). Plant extracts were prepared by Soxhlet extraction using 50% ethanol, followed by concentration with rotary evaporation and lyophilization. The extracts were combined in a 7:3 ratio (BB:BR) and fermented with *Lactobacillus plantarum* 2656 under controlled conditions to obtain a fermented product (FBBBR) with 60% yield. Antimicrobial activity of the formulation was assessed against *Shigella boydii* ATC 9207 using agar cup diffusion, resazurin-based minimum inhibitory concentration (MIC), and minimum bactericidal concentration (MBC) assays, confirming its inhibitory potential. Additionally, an orally disintegrating film (ODF) was formulated using carboxymethyl cellulose, gelatine, starch, and sorbitol as film-forming agents for the incorporation of the probiotic. The ODF was evaluated for swelling index, disintegration time in water and simulated saliva, pH, and probiotic viability during storage. Results demonstrated that the fermented extracts enhanced antimicrobial activity and maintained probiotic viability within the ODF matrix. Overall, this study highlights a novel formulation strategy that integrates natural bioactive extracts with probiotics, offering a promising functional nutraceutical delivery system with potential therapeutic benefits.

Keywords: Probiotics, fermentation, blueberry, black rice, *Lactobacillus plantarum*, oral disintegrating film, antimicrobial activity.

Introduction

Probiotics are defined as “live microorganisms which, when administered in adequate

*Corresponding Author Email: ap826893@gmail.com

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amounts, confer a health benefit on the host.” Their role in promoting gut health, preventing gastrointestinal disorders, and providing therapeutic benefits against infections has been widely studied. Probiotic strains exert their beneficial effects through multiple mechanisms, including competitive exclusion of pathogens, production of antimicrobial metabolites, modulation of the host immune system, and inhibition of toxin expression.

Traditionally, probiotics are consumed through fermented foods such as yogurt, cheese, kimchi, sauerkraut, kombucha, and miso, as well as dietary supplements in capsule or powder form. However, many commercially available probiotics face challenges such as poor survival during gastrointestinal transit and reduced viability during storage. To overcome these limitations, advanced formulation techniques such as spray-drying, freeze-drying, fluidized bed drying, and extrusion have been developed to enhance stability and targeted delivery of probiotic strains.

The therapeutic applications of probiotics are diverse, ranging from the prevention of antibiotic-associated and traveller’s diarrhoea to reduction of respiratory infections, mitigation of irritable bowel syndrome, alleviation of atopic dermatitis, and support in eradicating *Helicobacter pylori*. They have also shown potential in managing inflammatory bowel diseases and other systemic conditions, underscoring their significance in both preventive and therapeutic healthcare.

In recent years, functional foods combining probiotics with natural bioactive compounds have gained increasing attention. Black rice (*Oryza sativa L. indica*), rich in anthocyanins, fibers, and micronutrients, has demonstrated antioxidant, anti-inflammatory, and anti-diabetic properties. Similarly, blueberries (*Vaccinium corymbosum*), abundant in flavonoids, phenolic acids, and vitamins, are known for their prebiotic, antimicrobial, antioxidant, and cardioprotective activities. Together, these natural sources provide an excellent nutritional base to enhance the growth and efficacy of probiotic strains.

Among probiotics, *Lactobacillus plantarum* is one of the most versatile species, naturally found in fermented foods and the human gastrointestinal tract. It produces antimicrobial substances effective against both Gram-positive and Gram-negative pathogens, thereby contributing to gut microbiota balance and protection against infections. Notably, pathogens such as *Shigella boydii*, a causative agent of bacillary dysentery, remain a major concern in developing regions, highlighting the need for novel probiotic-based strategies for prevention and treatment.

The present study aims to develop and formulate a probiotic using a fermentation-based approach, incorporating black rice and blueberry extracts as natural substrates. The study further evaluates the antimicrobial activity of the formulation and explores the potential of incorporating the probiotic into an orally disintegrating film (ODF) for enhanced delivery and stability. This innovative strategy integrates natural bioactive compounds with probiotic technology to create a functional nutraceutical system with significant therapeutic potential.

Materials and Methods

2.1 Extraction and Purification of Plant Materials

Black rice (*Oryza sativa L. indica*) and blueberry (*Vaccinium corymbosum*) were procured from local certified suppliers. The raw materials were dried, powdered, and subjected to Soxhlet extraction using 50% ethanol as solvent. Extraction was carried out for 24 h at 150 °C, followed by concentration using a rotary vacuum evaporator (BR: 90–120 rpm; BB: 80–140 rpm). The dried extracts were freeze-dried at –80 °C and stored in airtight containers. The extraction yield was 8% for black rice and 12% for blueberry.

2.2 Microorganisms

The probiotic strain *Lactobacillus plantarum* 2656 was obtained from NCIM, CSIR-NCL, Pune, India. For antibacterial assays, *Shigella boydii* ATC 9207 was used as the test pathogen. Additional probiotics (*L. acidophilus* Ki, *L. rhamnosus* R11, *Bifidobacterium animalis* Bb12, and *B. animalis* Bo) and pathogens (*Escherichia coli* NCTC 9001, *Salmonella enteritidis* ATCC 13076, *Listeria monocytogenes* ESB 3562, and *Bacillus cereus* NCTC 2599) were included for comparative evaluation as per standard culture collection protocols.

2.3 Fermentation of Extracts

The blueberry and black rice extracts were mixed in a 7:3 ratio (BB:BR) and inoculated with *L. plantarum* 2656. Fermentation was carried out in a shaking incubator at 37 °C and 120 rpm for 24 h. The fermented product (FBBBR) was freeze-dried and stored at –20 °C. The overall yield of the fermented extract was approximately 60%.

2.4 Antibacterial Activity

2.4.1 Agar Cup Diffusion Assay

The antimicrobial activity of FBBBR against *S. boydii* was assessed using the agar cup diffusion method. Test cultures were grown in Mueller–Hinton broth for 24 h, adjusted to 0.5 McFarland standard ($\sim 1 \times 10^8$ CFU/mL), and further diluted to $\sim 1 \times 10^6$ CFU/mL. Wells containing 100 μ L of the fermented extract were loaded, with chloramphenicol and sterile distilled water serving as controls. Plates were incubated at 37 °C for 24 h, and zones of inhibition were recorded.

2.4.2 Minimum Inhibitory Concentration (MIC)

The MIC of the fermented extracts was determined using the Resazurin Microtiter Assay (REMA). Serial two-fold dilutions of samples were prepared in 96-well plates containing Mueller–Hinton broth. Each well was inoculated with 1×10^4 CFU/mL of *S. boydii* and incubated at 37 °C for 24 h. Resazurin (10 μ L of 2 mg/mL) was added to each well, and color change was monitored after 30 min. The MIC was defined as the lowest concentration showing no color change.

2.4.3 Minimum Bactericidal Concentration (MBC)

Aliquots from wells corresponding to the MIC and higher concentrations were sub-cultured onto Mueller–Hinton agar plates. Plates were incubated at 37 °C for 24 h. The MBC was defined as the lowest concentration resulting in $\geq 99.9\%$ reduction in viable counts compared to the initial inoculum.

2.5 Development of Oral Disintegrating Films (ODF)

ODFs were prepared by the solvent casting method using carboxymethyl cellulose (CMC), gelatin, starch (0.5:0.5:0.5), and sorbitol as plasticizer. Solutions were prepared separately, hydrated, heated, and combined, followed by addition of probiotic culture. The homogenized solution was degassed by ultrasonication and poured into plates (8×9 cm). After drying for 24 h at room temperature, the films were cut into uniform pieces and stored under vacuum and non-vacuum conditions.

2.6 Evaluation of ODFs

- **Swelling Index:** Measured by immersing the film in 7 mL of distilled water and recording weight gain at 30 s intervals.
- **Disintegration Time:** Determined in distilled water and simulated saliva medium at 37 °C until complete breakdown of the film.
- **pH Measurement:** The film was dissolved in phosphate buffer (pH 7.45), and changes in pH were noted.
- **Probiotic Viability:** Films were stored at 20 °C under vacuum and non-vacuum conditions. Viability was assessed periodically by plating on MRS agar and determining colony-forming units (CFU).

Results

- Extraction of blueberry (BB) and black rice (BR) yielded total residues of 1,232 ml and 1,190 ml, respectively. After rotary evaporation, the concentrated extracts provided 12% residue for BB and 8% for BR. The final fermentation product volume obtained was 20 ml.
- The antibacterial activity of the BB:BR (7:3) formulation was assessed against *Shigella boydii* ATCC 9207. Zone of inhibition measurements revealed activity proportional to the concentration used, with 22 nm at 50 μ l and 26 nm at 100 μ l, compared to the positive control (40 nm). Distilled water showed no inhibitory activity.



Figure No 1: Sample against *Shigella boydii*

Table No1. Zone of Inhibition against *Shigella boydii*

Sr. no.	Organism	Sample Concentration &	Zone of Inhibition (nm)
1	<i>Shigella boydii</i> ATC 9207	B+R 50 µl	22
		B+R 100 µl	26
		PC	40
		DW	-

- The minimum inhibitory concentration (MIC) of the formulation against *S. boydii* was consistently observed at a 1:6 dilution, while the minimum bactericidal concentration (MBC) was confirmed within the range of 1:4 to 1:7 dilutions.

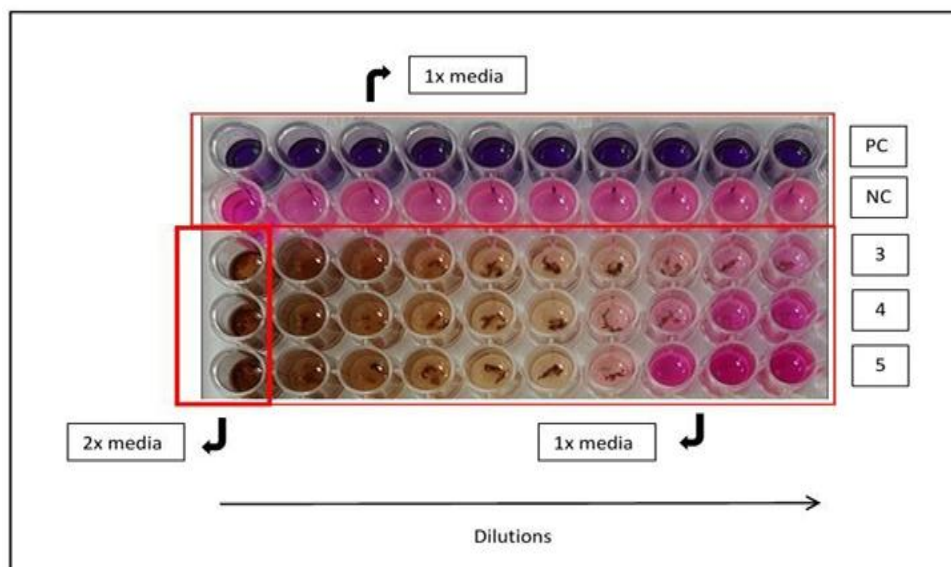


Figure No 2: Resazurin microtiter plate minimum inhibitory concentrations

Table no 2: Dilutions of the Samples

Sr. no.	Culture	Conc. 1	Conc. 2	Conc. 3	Conc. 4	Conc. 5	Conc. 6	Conc. 7	Conc. 8	Conc. 9
1	PC	PC	PC	PC	PC	PC	PC	PC	PC	PC
2	DW	DW	DW	DW	DW	DW	DW	DW	DW	DW
3	BB and BR	1:1	1:2	1:3	1:4	1:5	1:6	1:7	1:8	1:9
4	BB and BR	1:1	1:2	1:3	1:4	1:5	1:6	1:7	1:8	1:10

5	BB and BR	1:1	1:2	1:3	1:4	1:5	1:6	1:7	1:8	1:10
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Table No 3: Observed Minimum Inhibitory Concentrations

Sr. no.	Organism	Minimum Inhibitory Concentration (Dilution)
1	Shigella boydii	1:6

Minimum bactericidal concentrations

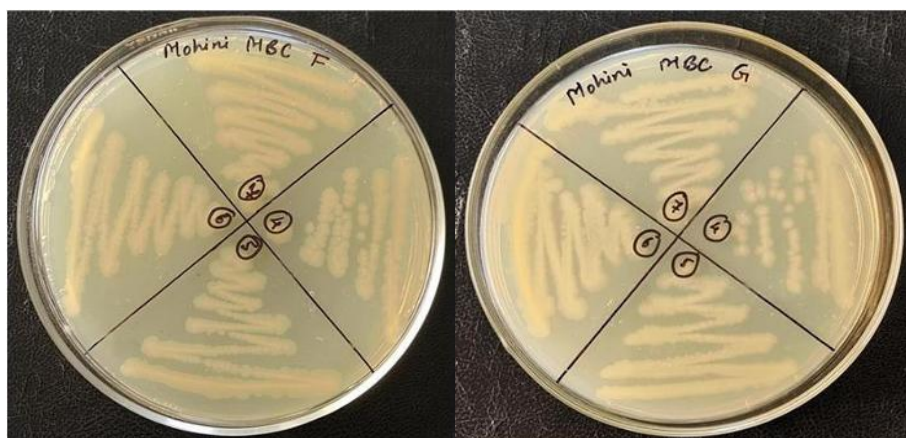


Figure no 3 Plate 1: MBC results for 1:4, 1:5, 1:6 & 1:7 dilution of the 3th row

Figure no4 Plate 2 : MBC results for 1:4, 1:5, 1:6, & 1:7 dilutions of the 4th row

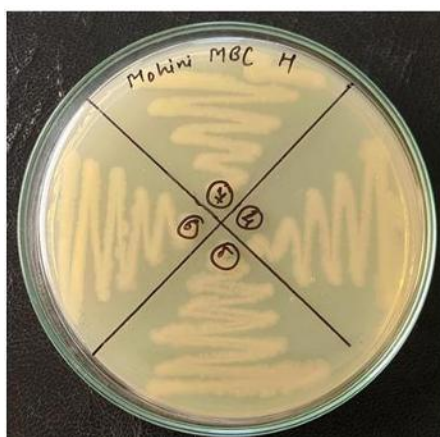


Figure no 5 Plate 3 : MBC results for 1:4, 1:5, 1:6 & 1:7 dilutions of the 5th row

- Physicochemical characterization of the oral dispersible film (ODF) formulation showed promising results. The swelling test indicated a gradual increase in weight from 0.242 g initially to 0.274 g after 90 seconds, confirming hydration capacity. The

disintegration test (DT) demonstrated dissolution of the ODF within 3 minutes in distilled water and 6 minutes in simulated saliva, both within the pharmacopeial standards (2.5–6.5 min).

- The pH of the product was measured at 7.2, aligning with the physiological salivary pH range (6.9–7.4).

Discussion

This study demonstrates the successful development of a probiotic formulation combining blueberry (*Vaccinium corymbosum*) and black rice (*Oryza sativa* L. indica) extracts fermented with *Lactobacillus plantarum* 2656. The fermentation process yielded a stable product with enhanced antimicrobial potential, supporting the hypothesis that polyphenol-rich plant substrates can improve probiotic activity.

The antimicrobial assays confirmed the inhibitory effect of the formulation against *Shigella boydii*, an enteric pathogen of global health concern. The observed inhibition zones (22–26 mm) were smaller than the positive control but indicate a measurable antimicrobial effect attributable to synergistic action between phytochemicals and probiotic metabolites. The MIC (1:6 dilution) and MBC (1:4–1:7 dilutions) values reinforce the bacteriostatic and bactericidal efficacy of the formulation at relatively low concentrations, suggesting practical therapeutic potential. Previous studies have similarly reported that anthocyanin- and flavonoid-rich substrates enhance lactic acid bacteria activity by providing growth-promoting phenolic compounds and generating antimicrobial metabolites during fermentation.

The oral dispersible film (ODF) developed as a delivery system exhibited desirable pharmaceutical characteristics, including rapid disintegration (3 min in water, 6 min in saliva) and acceptable pH (7.2), which ensures compatibility with oral physiology. The swelling index confirmed hydration capacity, essential for rapid release and bioavailability. Such properties make ODFs a patient-friendly dosage form, particularly for pediatric and geriatric populations, overcoming limitations of conventional probiotic supplements such as capsules and powders.

Collectively, these findings suggest that the integration of natural extracts with probiotic technology enhances both functionality and stability. While the study focused primarily on *S. boydii*, broader antimicrobial screening against other clinically relevant pathogens would provide a more comprehensive evaluation. Additionally, in vivo studies and stability assessments over extended storage are needed to validate clinical applicability.

Conclusion

The present study successfully formulated a probiotic product by fermenting blueberry and black rice extracts with *Lactobacillus plantarum* 2656 and incorporating it into an oral dispersible film. The formulation demonstrated notable antimicrobial activity against *Shigella boydii*, with favourable MIC and MBC values, along with optimal physicochemical properties such as rapid disintegration and physiological pH compatibility. These results highlight the potential of combining plant bio actives with probiotics in innovative delivery systems to enhance therapeutic efficacy and consumer compliance.

This work contributes to the growing field of functional nutraceuticals, offering a novel approach to probiotic delivery. Future investigations should focus on long-term stability, large-scale production feasibility, and clinical trials to establish efficacy and safety in human populations.

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